

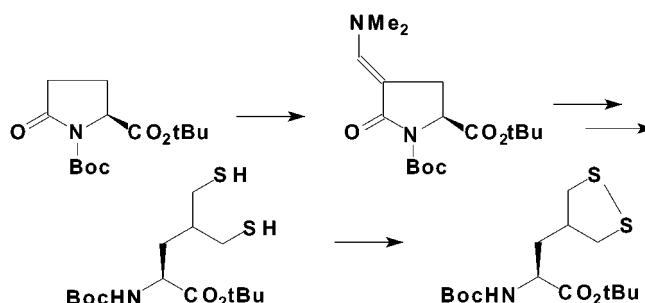
Synthesis of 1,2-Dithiolane Analogues of Leucine for Potential Use in Peptide Chemistry

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ABSTRACT



1,2-Dithiolanes present several points of interest for both peptide and medicinal chemistry, yet no chiral α-amino acids containing this five-membered heterocyclic system are available. We report here the first synthesis of *N*- and *C*-protected derivative of (*S*)-2-amino-3-(1,2-dithiolan-4-yl)propionic acid (Adp) and its 1,3-dithiolic form.

Proteinogenic α-amino acids represent an exceptionally rich source of chiral building blocks and synthons to be used in both organic and medicinal chemistry.¹ Nonproteinogenic and synthetic amino acids are, however, becoming increasingly important for the design of peptidomimetics endowed with improved specific properties.² It is well-known in fact that, although native biologically active peptides have a great potential as therapeutic agents, they need to be modified to overcome problems connected with metabolic stability, appropriate bioavailability, and receptor selectivity.

Among nonproteinogenic α-amino acids, particular attention is being devoted to models whose side chain bears a heterocyclic ring system;³ these compounds present in fact

a variety of chemical structures and, when appropriately inserted into peptide backbones, offer the opportunity to establish efficient interactions with the biomolecular targets.

In this context, the heterocyclic five-membered ring of 1,2-dithiolanes should represent an interesting molecular fragment to be introduced into α-amino acid side chains. Due to the stereoelectronic effects involving the sulfur lone pairs and the small dihedral angle around the S–S bond,^{4,5} the chemical and physical properties of 1,2-dithiolanes, which represent the oxidized form of 1,3-dithiols, are very distinct from those of open chain and large ring disulfides; of particular relevance, in the field of peptidomimetics, are the fast rate of ring opening by nucleophiles,^{6,7} the activity as

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antioxidant and free-radical scavengers,⁸ and the efficient metal-ion coordinating properties.⁹ It should be remembered here that the 1,2-dithiolane ring system represents the key structural feature on which the reactivity of relevant biomolecules such as that of lipoic acid (**1**, Figure 1), the

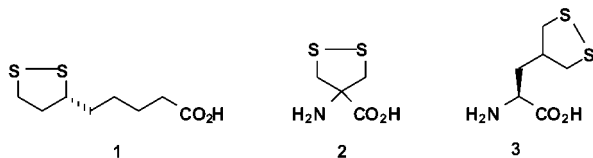


Figure 1. Structure of 1,2-dithiolane derivatives: naturally occurring *R*-enantiomer of 1,2-dithiolane-3-pentanoic acid (lipoic acid, **1**); 4-amino-1,2-dithiolan-4-carboxylic acid (Adt, **2**); (*S*)-2-amino-3-(1,2-dithiolan-4-yl)propionic acid (Adp, **3**).

essential cofactor in the oxidative decarboxylation of α -ketoacids, is based.^{9,10}

On the basis of the above-reported considerations and by considering the absence of information on the chemistry of peptidomimetics incorporating 1,2-dithiolane heterocyclics, we recently began a research program centered on the study of this topic. The $C^{\alpha,\alpha}$ -tetrasubstituted and achiral residue of 4-amino-1,2-dithiolane-4-carboxylic acid (Adt, **2**) was chosen as an initial model and the conformational and biochemical consequences of its insertion into bioactive oligopeptides have been described.^{11,12} As a continuation of these studies we report here the stereocontrolled synthesis of the *N*- and *C*-protected derivatives of (*S*)-2-amino-3-(1,2-dithiolan-4-yl)propionic acid (Adp, **3**) and its 1,3-dithiolic form **14**. The Adp molecule represents the first example of chiral an α -amino acid containing a cyclic disulfide in its side chain.

The use of the malonic precursor, obtained from (*S*)-pyroglutamic acid or (*S*)-serine derivatives, as depicted in Scheme 1, was initially considered a suitable approach to prepare **3**. However, the regioselective reduction of this compound was found to be sluggish and gave complex reaction mixtures. Difficulties during the reduction of *N*-protected γ -carboxy glutamyl derivatives analogous to that reported in Scheme 1 have already been noted by Dubois et al.¹³ and are attributable, at least in part, to the participation of the urethane NH and formation of pyrrolidine derivatives.

(6) Concerning nucleophilic reactions at the sulfur atoms, it should be noted (see: Schmidt, U.; Grafen, P.; Goedde, H. W. *Angew. Chem., Int. Ed. Engl.* **1965**, *4*, 846) that the ground state of the 1,2-dithiolane ring system (valence angle at the S atom, 92°; S–S bond length, 2.1 Å) is much more similar geometrically to the transition state than is the ground state of an open chain disulfide (valence angle at S, 107°; S–S distance, 2.05 Å).

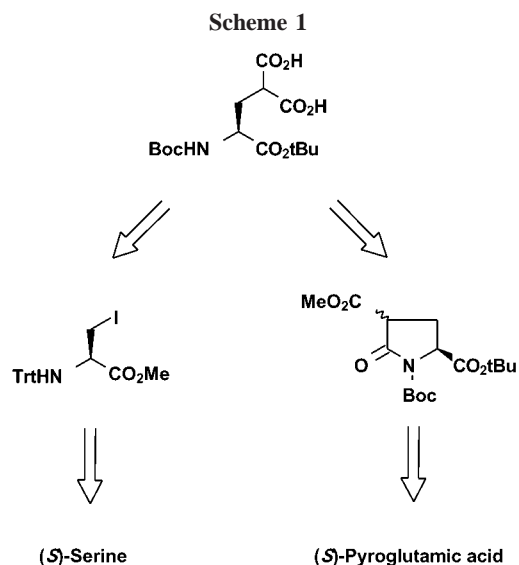
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An alternative approach (see Scheme 2), which follows in part the route previously used to prepare (*S*)-5,5'-dihydroxyleucine,¹⁴ was then adopted.

tert-Butyl (*S*)-*N*-*tert*-butoxycarbonylpyroglutamate **6** was used as the starting material.¹⁵ This was converted in high yield into the enaminone **7** by using *tert*-butoxy-bis(dimethylamino)methane (Bredereck's reagent).¹⁶ The NaBH₃CN reduction of the intermediate aldehyde **8**, obtained by acidic hydrolysis of **7**, at a pH value between 3.5 and 4.0, afforded a diastereoisomeric mixture of *cis* and *trans* alcohols **9** with improvement of the yield and shortening of the reaction time as compared with the original protocol.¹⁴ The mixture of the alcohols was then hydrolyzed using aqueous LiOH in THF, and the resulting hydroxy acid **10** was regioselectively reduced after conversion into the corresponding mixed anhydride and in situ treatment with NaBH₄. The synthesis of the dimesylate **12** was followed by treatment with potassium thiolacetate to give the bis-mercaptoacetyl derivative **13** in 85% combined yields. Aqueous alkaline hydrolysis of **13** at 0 °C afforded the (*S*)-5,5'-dimercaptoleucine derivative **14**¹⁷ whose iodine oxidation furnished

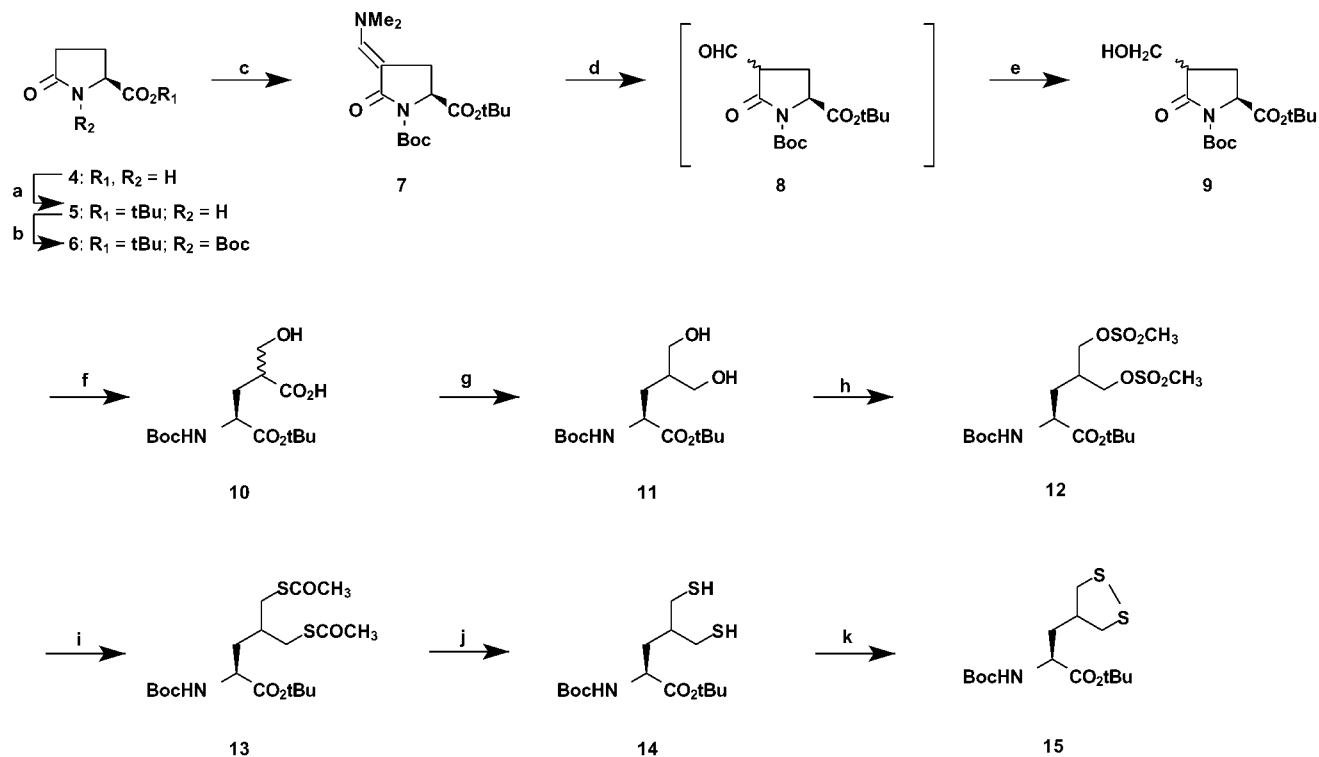
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(15) Protection as the *N*-Boc *tert*-butyl ester allows a regioselective hydrolysis during the ring-opening step and preserves the stereochemical integrity of the α -center (see: August, R. A.; Khan, J. A.; Moody, C. M.; Young, D. W. *Tetrahedron Lett.* **1992**, *33*, 4617).

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(17) (*S*)-*N*-Boc-5,5'-dimercaptoleucine *tert*-Butyl Ester (**14**). A solution of bis-thioacetate derivative **13** (0.135 g, 0.31 mmol) in degassed EtOH (2 mL) was treated with 0.93 mL of aqueous 1 N NaOH at 0 °C for 1 h. The mixture was neutralized with 2 N HCl and extracted with CH₂Cl₂. The organic phase, washed with water, dried, and evaporated at room temperature, gave an oily residue of pure **14** (0.108 g, 99%). [α]_D⁺⁹ (*c* = 2.0; CHCl₃); IR (CHCl₃) 3434, 2981, 1706, 1501, 1369, 1236, 1153 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 1.24 (m, 1H, SH_A), 1.32 (t, 1H, *J* = 8.5 Hz, SH_B), 1.44 and 1.47 (2 × s, 18H, 2 × *tert*-butyl), 1.65 (m, 1H, β -CH_A), 1.87 (m, 2H, β -CH_B and γ -CH), 2.70 and 2.82 (2 × m, 4H, 2 × CH₂S),

Scheme 2^a

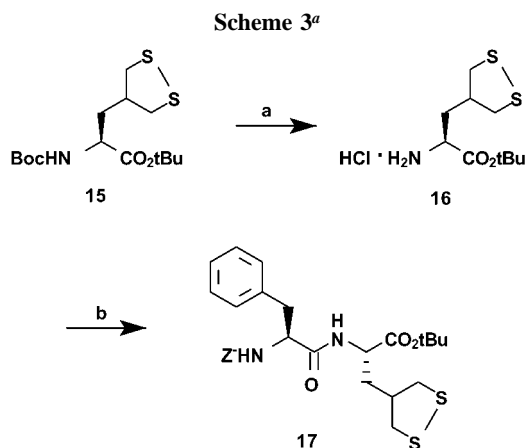
^a HClO₄ 70%, CH₃CO₂tBu, 76%; (b) (Boc)₂O, DMAP, 94%; (c) Bredereck's reagent, DME, 75 °C, 88%; (d) aqueous 0.2 N HCl, MeOH; (e) NaBH₃CN, 4 h, 56% from **7**; (f) aqueous 1 M LiOH, THF, 0 °C, 95%; (g) *i*BuOCOC₂H₅, NMM, THF, -15 °C, then NaBH₄ at 0 °C, 79%; (h) CH₃SO₂Cl, TEA, CH₂Cl₂ dry, 0 °C, 94%; (i) CH₃COSK, DMF, 91%; (j) aqueous 1 N NaOH, EtOH, 0 °C, 99%; (k) CHCl₃, AcONa, aqueous 0.1 M I₂, 98%.

eventually the desired Adp derivative **15**¹⁸ in almost quantitative yield; this latter compound was also obtained by a *one-pot* procedure starting from **13** (72%). As expected, and in accordance with the chemistry of 1,2-dithiolanes,⁵ the new α -amino acid derivative **15** can be easily reduced to regenerate the 5,5'-dimercaptolucine **14**.

4.15 (m, 1H, α -CH), 5.09 (d, 1H, $J = 8.4$ Hz, NH); ¹³C NMR (75.43 MHz, CDCl₃) δ 26.3 and 27.0 (2 \times CH₂S), 27.9 and 28.2 [2 \times C(CH₃)₃], 35.0 (β -CH₂), 39.5 (γ -CH), 51.8 (α -CH), 79.7 and 82.0 [2 \times C(CH₃)₃] 155.1 and 171.4 (2 \times CO).

(18) *N*-Boc-Adp *tert*-Butyl Ester (**15**). An aqueous solution of 0.1 M iodine was added dropwise at 0 °C to a solution of **14** (0.096 g, 0.273 mmol) and AcONa (0.067 g, 0.820 mmol) in CHCl₃ (8 mL) until a persistent pale yellow color appeared (2.9 mL). The mixture was decolorized with 1 M Na₂S₂O₃ and the organic phase was separated, washed with water, dried, and evaporated at room temperature. The crude (0.101 g) was chromatographed on silica gel (4 g) using CHCl₃ as eluent to give 0.093 g of **15** (98%). Mp 61–61.5 °C (hexane); [α]_D +21° ($c = 1.0$; CHCl₃); IR (CHCl₃) 3434, 2982, 1707, 1500, 1369, 1153 cm⁻¹; UV (CHCl₃) λ_{max} 332 (152); ¹H NMR (300 MHz, CDCl₃) δ 1.44 and 1.47 (2 \times s, 18H, 2 \times *tert*-butyl), 1.71 (m, 1H, β -CH_A), 1.97 (m, 1H, β -CH_B) 2.68 (m, 1H, γ -CH), 2.86 and 3.31 (2 \times m, 4H, 2 \times CH₂S), 4.19 (m, 1H, α -CH), 5.10 (d, 1H, $J = 8.1$ Hz, NH); ¹³C NMR (75.43 MHz, CDCl₃) δ 27.9 and 28.3 [2 \times C(CH₃)₃], 37.3 (β -CH₂), 43.7 and 44.0 (2 \times CH₂-S), 44.1 (γ -CH), 53.1 (α -CH), 79.8 and 82.2 [2 \times C(CH₃)₃], 155.1 and 171.2 (2 \times CO). The Adp derivative **15** could also be prepared by a *one-pot* procedure: thus, a solution of bis-dimercaptoacetyl derivative **13** (0.519 g, 1.19 mmol) in EtOH (8 mL) was treated with 3.56 mL of aqueous 1 N NaOH at 0 °C for 1 h. The mixture was diluted with CH₂Cl₂ (60 mL) and then an aqueous solution of 0.1 M iodine was added dropwise until a persistent pale yellow color appeared. The mixture was decolorized with 1 M Na₂S₂O₃ and the organic phase was separated, washed with water, dried, and evaporated at room temperature. The crude (0.385 g) was chromatographed on silica gel (12 g) using CHCl₃ as eluent to give 0.299 g of **15** (72%).

Although the Adp derivative **15** contains two *tert*-butyl protecting groups, it was possible, by adopting the procedure described by Rapoport et al.,¹⁹ to selectively remove the *N*-Boc protection in the presence of the *tert*-butyl ester; Scheme 3 reports an application of this procedure together with the synthesis of the dipeptide *Z*-Phe-Adp-*O*tBu.



^a 1 M HCl in EtOAc, rt, 4 h, 69%; (b) HOBt, EDC, *Z*-Phe-OH, TEA, DMF, 79%.

In conclusion, we have described the efficient synthesis of a pair of chiral α -amino acids structurally related to the natural residue of leucine and characterized by the presence in its side chain of the 1,2-dithiolane-1,3 dithiol redox system. By considering the relevant biological role played by the lipoic-dihydrolipoic acid system as well as the

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peculiar reactivity of 1,2-dithiolanes, the incorporation of these new residues into bioactive peptides should represent a new and promising approach.

Supporting Information Available: Experimental procedures and characterization data for compounds **12**, **13**, **16**, and **17**. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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